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(Maciel et al. 2003. *Comp. Biochem. Physiol. B* 134:641–649). In Australia, where *R. marina* was introduced, *Crocodylus johnstoni* individuals of intermediate size class died after the ingestion of the toads due to the absence of physiological adaptations against its highly toxic secretions (Letnic et al. 2008. *Biol. Conserv.* 141:1773–1782). However, *Crocodylus porosus*, which naturally coexist with *R. marina*, presented a better performance against the bufotoxin when compared to *C. johnstoni* during physiological tests (Smith and Phillips 2008. *Pac. Conserv. Biol.* 12:40–49). Predation behavior on *Rhinella marina* by Schneider's Dwarf Caiman (*Paleosuchus trigonatus*) was described in the state of Pará, Brazil (Assis and Santos 2007. *Herpetol. Rev.* 38:45); however, there are no data available regarding the interaction between *R. marina* and *C. crocodilus*. Among terrestrial vertebrates reported in the diet of *C. crocodilus*, anurans are few (Magnusson et al. 1987. *J. Herpetol.* 21:85–95; Thorbjarnarson 1993, *op. cit.*; Da Silveira and Magnusson 1999. *J. Herpetol.* 33:181–192). Percentage of prey occurrence analysis and relative digestibility can bias the presence of vertebrate prey items in the diet of *C. crocodilus* (Thorbjarnarson 1993, *op. cit.*). We believe that field observations on crocodylian foraging behavior can provide relevant data about predator-prey interactions. Meanwhile, captive experiments can help in understanding feeding habits and preferences as well as in investigating resistance to bufotoxins.

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CROCODYLUS MORELETTI (Morelet's Crocodile). SMALL POND USE. At 0047 h on 14 June 2008, during rainy conditions, we encountered a juvenile (571 mm SVL) female *Crocodylus moreletii* in a small pond (3.0 m × 4.3 m, 70 mm deep) in the middle of a small road surrounded by a brackish water wetland in Reserva de la Biosfera Los Petenes, Campeche, México (20.087032°N, 90.401258°W). At 0139 h, we observed another juvenile female *C. moreletii* (700 mm SVL) in a second small pond (1.4 × 3.0 m, 90 mm deep at 20.089948°N, 90.433983°W).

These observations are of interest since both ponds were in the middle of a small road surrounded by brackish water wetlands that are more indicative habitat for this species. We encountered another three *C. moreletii* (one adult, two juveniles) in open waters within the actual wetlands the same night. No previous reports exist of this behavior (Alvarez del Toro and Sigler 2001. *Los Crocodylia de México*. IMERNAR, PROFEPA, México 134 pp.). It is possible that the animals utilizing these smaller sites may take advantage of the low salinity in the rainwater filled ponds or the small ponds may provide food resources based on the potential prey we observed in similar ponds on this area (*Lithobates brownorum*, *Leptodactylus melanonotus*, *Ollotis valliceps*, *Smilisca baudinii*).

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SQUAMATA — LIZARDS

AMEIVA CORAX (Little Scrub Island Ground Lizard). FEEDING BEHAVIOR. *Ameiva corax* is endemic to Little Scrub Island, off the northeast coast of Anguilla in the British West Indies (18.308°N, 62.967°W). It is reported to consume the eggs of seabirds that nest on the island (Hodge et al. 2003. *The Reptiles and Amphibians of Anguilla, British West Indies*. Anguilla National Trust, Anguilla. 72 pp.). Herein we report on an individual lizard of this species opening, partially consuming, and ultimately killing the chick of a Laughing Gull (*Larus atricilla*) nesting on the island (Fig. 1).

At 0904 h on 12 June 2009 we noticed a lizard in a gull nest attempting to open one of three eggs present. The egg was opened in less than one minute; it is unclear whether the lizard cracked the egg or whether it had already been cracked by an emerging chick. We have observed other lizards cracking and consuming the contents of gull eggs in earlier stages of development, but since it appeared to be near hatching, the chick may have cracked the egg itself. Nevertheless, the lizard successfully opened the egg and we monitored its activity from ca. 5 m away. Once the egg was opened, the lizard first consumed the yolk sac, then began to claw and bite at the lower abdomen of the chick, eventually opening a hole in the chick's skin, pulling out and consuming part of its intestinal tract. Although the lizard bit and clawed at other parts of the chick, particularly the legs and wings, it did not pierce the skin elsewhere. The lizard (individually identifiable by



FIG. 1. *Ameiva corax* consuming the intestinal tract of a Laughing Gull (*Larus atricilla*) chick.

its shedding skin pattern) left the nest and returned three times between 0910 and 0930 h. When the lizard left the nest at 0930 h, the chick was still alive and moving. An adult gull returned to the nest at 0930h and incubated the remaining eggs. We monitored the nest until 1000 h, at which point the chick was still alive and an adult gull was on the nest. The chick was dead at 1230 h when we next checked on it.

We thank the Government of Anguilla for allowing us to work on Little Scrub Island. We are particularly grateful to Karim Hodge and Rhon Conner in the Department of the Environment for their support, encouragement, and advice. We also thank Captain Nature Boy Webster for transporting us to and from the island every day.

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AMEIVA LEPTOPHRYS (Delicate Ameiva). ENDOPARASITES. *Ameiva leptophrys* frequents lowland humid forests of southwestern Costa Rica and Panama to western Colombia (Savage 2002. The Amphibians and Reptiles of Costa Rica. A Herpetofauna Between Two Continents, Between Two Seas. Univ. Chicago Press, Chicago, Illinois. 934 pp.). There are, to our knowledge, no reports of helminths from *A. leptophrys*. The purpose of this note is to establish the initial helminth list for *A. leptophrys* as part of an ongoing survey of lizard helminths from Central America.

Six *A. leptophrys* (mean SVL = 112.3 mm \pm 17.4 SD, range: 85–130 mm) from Costa Rica were examined from the herpetology collection of the Natural History Museum of Los Angeles County (LACM), Los Angeles, California: Limón Province (LACM 174920); Puntarenas Province (LACM 174922–174924, 174927, 174928). Lizards were collected 1963–1967. The digestive tract was removed and the esophagus, stomach, small and large intestines were examined for helminths using a dissecting microscope. Nematodes and cystacanths were cleared in a drop of glycerol on a microscope slide, cover-slipped and studied using a compound microscope. Three species of Nematoda were found: *Physaloptera retusa* (stomach) prevalence, number infected/number examined \times 100 = 83%, mean intensity, mean number helminths \pm 1 SD = 49.8 \pm 33.6 SD, range = 2–85; *Spinicauda spinicauda* (large intestine) prevalence = 67%, mean intensity = 3.8 \pm 2.2 SD, range = 1–6 and two ascarid larvae (Angusticaecinae) (body cavity) prevalence = 17% and one species of Acanthocephala as 11 cystacanths (body cavity) prevalence = 17%. Helminths were deposited in the United States National Parasite Collection (USNPC), Beltsville, Maryland: *Physaloptera retusa* (USNPC 103860); *Spinicauda spinicauda* (USNPC 103861); Ascarid larvae (USNPC 103862); Oligacanthorhynchid cystacanths (USNPC 103863).

Physaloptera retusa is widely distributed in the Western Hemisphere. Records are summarized in McAllister et al. (2010. Comp. Parasitol. 77:184–201). Physalopterid nematodes utilize insect intermediate hosts (Anderson 2000. Nematode Parasites of Vertebrates. Their Development and Transmission. CABI Publishing, Oxon, UK, 650 pp.). Lizards are infected through their diet. *Spinicauda spinicauda* is known from a variety of South American lizards (Baker 1987. Mem. Univ. Newfoundland, Misc. Pap. 11:1–325; Ramallo et al. 2009. J. Parasitol. 95:1026–1028). Lizards become infected when they ingest eggs; no intermediate host is utilized (Anderson 2000, *op. cit.*). Eggs of various species assigned to the order Ascaridida, Angusticaecinae will hatch in a

variety of vertebrate intermediate hosts and develop into third-stage infective larvae (Anderson 2000, *op. cit.*). Because these larvae were in cysts, the possibility that *A. leptophrys* was serving as a transport (= paratenic host) must be considered. Cystacanths are larval forms infective to the definitive host (Kennedy 2006. Ecology of the Acanthocephala. Cambridge Univ. Press, Cambridge, UK. 249 pp.) and reach the lizard when it ingests an infected insect. No further development occurs in the lizard which serves as a paratenic transport host. *Physaloptera retusa*, *Spinicauda spinicauda*, ascarid larvae and oligacanthorhynchid cystacanths in *A. leptophrys* are new host records. Costa Rica is a new locality record for *S. spinicauda*.

We thank Christine Thacker (LACM) for permission to examine individuals of *A. leptophrys*, which are part of the Costa Rica Expeditions Collection donated to LACM in 1998 by Jay M. Savage. Cecilia Nava and Daisy Salguero (Whittier College) assisted with dissections.

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ANOLIS ALTAE (High Anole). ENDOPARASITES. *Anolis altae* occurs in premontane and lower montane zones of the Cordillera de Tilarán, Cordillera Central and Cordillera de Talamanca of Costa Rica at elevations of 1220 to 2000 m (Savage 2002. The Amphibians and Reptiles of Costa Rica: A Herpetofauna Between Two Continents, Between Two Seas. Univ. Chicago Press, Chicago, Illinois. 934 pp.). Five *A. altae* (mean SVL = 45.2 mm \pm 1.6 SD, range: 44–48 mm) collected November 1963 and deposited in the herpetology collection of the Natural History Museum of Los Angeles County (LACM), Los Angeles, California, USA were examined for helminths (Costa Rica: Cartago Province, LACM 149056, 149063–149065, 169075).

The digestive tract was removed through a mid-ventral incision, and the esophagus, stomach and small and large intestines were examined for helminths under a dissecting microscope. Cestodes were found in the small intestine, nematodes were found in the small and large intestines. Cestodes were regressively stained in hematoxylin, dehydrated in ethanol and mounted in Canada balsam; nematodes were cleared in glycerol and cover-slipped on a microscope slide. Both preparations were studied under a compound microscope. The cestodes were identified as *Mathevotaenia panamaensis* (prevalence [number infected lizards/number lizards examined \times 100] = 40%, mean intensity [mean number helminths per infected lizard \pm 1 SD] = 36.0 \pm 39.6, range: 8–64). The nematodes were identified as *Falcaustra costaricae* (prevalence = 40%, mean = 1.0). Helminths were deposited in the United States National Parasite Collection (USNPC), Beltsville, Maryland: *Mathevotaenia panamaensis* (USNPC 103944); *Falcaustra costaricae* (USNPC 103945).

Mathevotaenia panamaensis was recently described from *Sceloporus malachiticus* from Panama by Bursley et al. (2010. Acta Parasitol. 55:53–57). *Anolis altae* is the second host to harbor this parasite although an unidentified species of *Mathevotaenia* was found in *Anolis humilis* from Nicaragua (Goldberg et al. 2010. Comp. Parasitol. 77:242–246). *Mathevotaenia panamaensis*, a member of the Anoplocephalidae, utilizes an arthropod as an intermediate host (Roberts and Janovy 2005. Gerald D. Schmidt